Absolute Stereochemistry of Spiciferones and Spicifernin, Bioactive Metabolites of the Fungus *Cochliobolus spicifer*: Evidence for their Unique Biosynthesis

Hiromitsu Nakajima,*,* Keiichi Fukuyama,^b Hiroaki Fujimoto,* Toshiyuki Baba*

and Takashi Hamasaki" ^a Department of Bio-resource Science, Faculty of Agriculture, Tottori University, Koyama, Tottori 680, Japan ^b Department of Biology, Faculty of Science, Osaka University, Toyonaka, Osaka 560, Japan

The absolute stereochemistry of spiciferone A and spicifernin, a phytotoxin and a plant-growth promoter, respectively, produced by the phytopathogenic fungus *Cochliobolus spicifer* Nelson, has been determined by the application of Mosher's method after establishment of relative stereochemistry by X-ray analysis. The absolute stereochemistry of spiciferones B and C has been determined by CD spectral comparison. Their absolute stereochemistry was in good agreement with the proposed polyketide biosynthetic pathway in which spiciferones and spicifernin are formed from a common cyclic precursor by a sequence of biological reactions involving a unique retro-aldol condensation.

Strain D-5 of *Cochliobolus spicifer* Nelson, which causes leaf spot disease in wheat, simultaneously produces several phytotoxins and a plant-growth promoter. The phytotoxins, *i.e.*, spiciferones A 1, B 2, and C 3 and spiciferinone 4, and the plantgrowth promoter, *i.e.*, spicifernin, tautomers 5a and 5b,† have been isolated and characterized in our laboratory.¹ Despite different carbon skeletons, they have unique structural features in common: (i) a quaternary carbon bearing an ethyl, a methyl, and a ketonic carbonyl and (ii) vicinal methyls. This commonality strongly suggests that these metabolites have the same origin. Labelled acetic acid and methionine were incorporated



into spiciferone A and spicifernin, and the resulting labelling patterns indicated the operation of the biosynthetic pathway such as in Scheme 1.² In this pathway a single hexaketide chain bearing two C-methyls from C_1 units is modified into spiciferone A and spicifernin via a 10-membered monocyclic intermediate 6 by a sequence of biological reactions including a unique retro-aldol condensation. In this hypothetical pathway the absolute configuration of C-8 in spiciferone A is closely related to that of C-7 in spicifernin through the intermediate 6. Thus their absolute configurations will afford evidence for this interesting biosynthetic pathway. However, those still remain to be determined. In this paper, we report the absolute

[†] Spicifernin was found to exist as an equilibrium mixture of tautomers **5a** and **5b** in solution from NMR data.^{1b}

stereochemistry of spiciferones and spicifernin and then discuss the validity of our proposed biosynthetic pathway.

Results and Discussion

The absolute stereochemistry of spiciferone A was determined by application of Mosher's method for its derivative whose relative stereochemistry was established by X-ray analysis. Spiciferone A 1 was converted into alcohols 7a and 7b in two steps as shown in Scheme 2 and these two alcohols were separable by HPLC as previously described.³ The major alcohol 7b, in which the hydroxy group is oriented in a pseudoaxial position, was found to be fairly nonreactive towards α -methoxy- α -(trifluoromethyl)phenylacetyl chloride (MTPACl) under ordinary conditions. Thus, the minor alcohol 7a was chosen for X-ray analysis and application of Mosher's method. The alcohol 7a was recrystallized from EtOAc-hexane to give needles and was subjected to X-ray analysis to establish its relative stereochemistry. The molecular structure of alcohol 7a is shown in Fig. 1 and atomic coordinates are listed in a supplementary publication.[‡] The relative configuration was found to be $(7R^*, 8S^*)$. The absolute configuration at C-7 of compound 7a was established by application of an advanced Mosher's method.⁴ Alcohol 7a was converted into its (R)- α methoxy- α -(trifluoromethyl)phenylacetic acid [(R)-MTPA] ester 8a and (S)-MTPA ester 8b, and their ¹H NMR spectra were measured in CDCl₃. The chemical-shift differences ($\Delta \delta$ = $\delta_{\rm sh} - \delta_{\rm sh}$) between the resonances in the ¹H NMR spectra of esters 8a and 8b are shown in Fig. 2. The absolute configuration at C-7 in the alcohol 7a was thus determined to be R by Mosher's method. Therefore, the absolute configuration at C-8 of compound 7a is S and consequently that of spiciferone A 1 was established to be R.

Next the CD spectra of spiciferones A, B and C, and the alcohol 7a were measured in order to establish the absolute stereochemistry of spiciferones B and C. Table 1 shows the CD spectral data for these compounds. The spectra of spiciferones revealed CD bands closely similar to each other, whereas the spectrum of alcohol 7a showed no perceptible CD bands at all. This indicates that the conjugated ketone functionality at C-7

[‡] For full details of the CCDC deposition scheme, see 'Instructions for Authors,' J. Chem. Soc., Perkin Trans. 1, 1994, issue 1.



Scheme 1 Proposed biosynthetic pathway² of spiciferone A 1, spiciferinone 4 and spiciferinin 5a/b. SAM = S-adenosylmethionine.

Table 1 CD spectral data for spiciferones A 1, B 2 and C 3, and the alcohol 7a

Compound	CD Band λ_{max} (EtOH)/nm ($\Delta \varepsilon$)
Spiciferone A 1	309 (-2.11) and 363 (+0.48)
Spiciferone B 2	305 (-1.62) and 360 (+0.35)
Spiciferone C 3	305 (-1.88) and 363 (+0.38)
Alcohol 7a	no perceptible CD band

is important for the characteristic CD bands of compounds 1-3 and also that C-8 in spiciferones B and C (compounds 2 and 3) has the same configuration as in spiciferone A 1.

The relative stereochemistry of spicifernin 5a/b was established by X-ray analysis and its absolute stereochemistry was determined by application of Mosher's method for the degraded product. Spicifernin was recrystallized from EtOAc-hexane to yield needles, which were subjected to X-ray analysis. The molecular structure of spicifernin is shown in Fig. 3 and atomic coordinates are listed in the supplementary publication. Thus, the relative stereochemistry of one form of spicifernin, 5a, was determined to be $(2S^*, 3R^*, 4R^*, 7S^*)$. Interestingly, spicifernin exists only as lactol 5a in the crystal state, while it shows acidic properties and exists as an equilibrium mixture of lactol 5a and keto acid 5b in solution.^{1b} Spicifernin was converted, via triester 9, into alcohol 10 in four steps, including Baeyer-Villiger oxidation, as shown in Scheme 2. Alcohol 10 was esterified with (+)- and (-)-MTPACl in pyridine to give (R)-MTPA ester 11a and (S)-MTPA ester 11b, respectively, and their ¹H NMR

spectra were measured in CDCl₃. The chemical-shift differences $(\Delta \delta = \delta_{11b} - \delta_{11a})$ between the resonances in ¹H NMR spectra of esters **11a** and **11b** are shown in Fig. 4. The absolute configuration for C-3 in the alcohol **10** was thus determined to be *R* by Mosher's method. The positive $\Delta \delta$ -value for methyl protons in the methoxycarbonyl function at C-7 is due to the following reason: this methyl is positioned on the same side of the MTPA plane as is the methyl at C-3 in the most preferable conformation of the MTPA esters **11a** and **11b**. This was confirmed by MM-2 conformational analysis of the corresponding (S)- α -methoxy- α -(methyl)phenylacetate † and (R)- α -methoxy- α -(methyl)phenylacetate † of alcohol **10** (Fig. 5).⁵ Thus, the absolute stereochemistry of one form of spicifernin, **5a**, was established as (2S,3R,4R,7S).

In our previous report,² we proposed the biosynthetic pathway shown in Scheme 1 for the production of spiciferone A and spicifernin in this fungus. In the proposed pathway they arise from the common monocyclic precursor 6 by retro-aldol condensation, which is a novel reaction in polyketide biosynthesis. If this pathway is operative in this fungus, the absolute configuration of the quaternary carbon in intermediate 6 is deduced to be S from the configuration of spiciferone A 1. Similarly, that carbon is deduced to have the S configuration from the stereochemistry of spicifernin. Therefore, our present results concerning the stereochemistry of spiciferone A and spicifernin strongly support the unique biosynthetic pathway we have proposed.

[†] *i.e.*, The corresponding α -methoxy- α -phenylpropionates.





Fig. 2 Determination of the absolute configuration at C-7 in alcohol 7a: $\Delta\delta$ -value was obtained from the ¹H NMR spectra of (*R*)-MTPA ester 8a and (*S*)-MTPA ester 8b



Fig. 3 Perspective view of spicifernin 5a drawn by ORTEP and the numbering system used for X-ray molecular structure



Scheme 2 Reactions of spiciferone A 1 and spicifernin 5b to establish their absolute stereochemistry. *Reagents and conditions:* i, H₂, PtO₂, EtOAc; ii, NaBH₄, THF, 20 min, 0 °C; iii, (*R*)- or (*S*)-MTPACl, pyridine, H₂NCH₂CH₂CH₂NMe₂; iv, CH₂N₂, Et₂O; v, MCPBA, NaH₂PO₄, CH₂Cl₂, 45 °C, 96 h; vi, Na₂CO₃, MeOH, room temperature.



Fig. 1 Perspective view of alcohol 7a drawn by ORTEP and the numbering system used for the X-ray molecular structure

Experimental

General Procedure.—Optical rotations were measured on a Horiba SEPA-200 high sensitivity polarimeter and $[\alpha]_{D}$ -values are given in 10^{-1} deg cm² g⁻¹. CD spectra were recorded on a



11a and 11b

Fig. 4 Determination of the absolute configuration at C-3 in alcohol 10: $\Delta\delta$ -value was obtained from the ¹H NMR spectra of (*R*)-MTPA ester 11a and (*S*)-MTPA ester 11b



Fig. 5 Stable conformation $(37.3 \text{ kcal mol}^{-1})$ of (S)- α -methoxy- α -phenylpropionate of alcohol 10 obtained by MM-2 energy minimization (1 cal = 4.184 J)

JASCO J-20C spectropolarimeter. MS were recorded on a JEOL DX-300 spectrometer. NMR spectra were measured on a JNM GX-270 FT NMR spectrometer (¹H, 270 MHz; ¹³C, 67.8 MHz), for solutions in CDCl₃ with Me₄Si as reference. The steric conformations were built up by MM-2 energy minimization of *CSC Chem3D*/*Plus*TM software (Cambridge Scientific Computing, Inc.) on a Macintosh Centris 650 computer.

X-Ray Crystal Structure Analysis.-Intensity data were

measured on a Rigaku four-circle diffractometer using Nifiltered Cu-K α radiation ($\lambda = 1.5418$ Å) and a rotating anode generator. The ω -2 θ scan mode was employed, with background measurement at the end of each scan. No significant change was observed in the intensities of the three standard reflections measured every 100 reflections for each compound. Intensity data were corrected only for Lorentz and polarization effects. Both structures were determined by direct methods using SHELX 86⁶ and refined by block-diagonal least-squares⁷ on F using atomic scattering factors.⁸ The hydrogen atoms were located in the difference Fourier maps. The oxygen and carbon atoms were refined anisotropically and the hydrogen atoms isotropically. Molecular structure perspective views (Figs. 1 and 3) were drawn by ORTEP-II.⁹

Crystal Data for Alcohol 7a.— $C_{14}H_{20}O_3$, M = 236.3, orthorhombic, space group $P2_12_12_1$, a = 12.637(2), b =7.552(1), c = 13.501(2) Å, V = 1288.5(3) Å³ (cell parameters by least-squares from the setting angles of 22 reflections), μ (Cu- K_{α} = 6.4 cm⁻¹, Z = 4, D_{X} = 1.22 g cm⁻³, F(000) = 512, T = 293 K, R = 0.050 and R_{W} = 0.062 for 1000 unique reflections with $F_0 > 2\sigma(F_0)$. A crystal of dimensions $0.05 \times 0.1 \times 0.6$ mm was used. Intensities of 1156 unique reflections were measured to $2\theta_{\max} = 120^{\circ}$ in the range $0 \le h \le 14, \ 0 \le k \le 8, \ 0 \le$ $l \le 15$, with ω scan width of $1.0^\circ + 0.15^\circ \tan \theta$, a scan speed of 4° min⁻¹, and a background counting time of 6 s. Strongest reflection, 0 2 0, was omitted from the refinement. The weighting scheme used in the final stage of refinement was $w = [\sigma(F_o)^2 +$ $0.015F_{o} + 0.0005F_{o}^{2}$]⁻¹. The residual electron densities in the final difference Fourier map ranged from -0.18 to 0.17 e Å⁻³ Atomic coordinates, temperature factors, and bond lengths and angles have been deposited at the Cambridge Crystallographic Data Centre.

Crystal Data for Spicifernin **5a**.—C₁₄H₂₂O₆, M = 286.3, orthorhombic, space group P2₁2₁2₁, a = 26.939(5), b =9.237(2), c = 5.995(1) Å, V = 1491.8(4) Å³ (cell parameters by least-squares from the setting angles of 21 reflections), μ (Cu-K α) = 8.7 cm⁻¹, Z = 4, $D_X = 1.27$ g cm⁻³, F(000) = 616, T =293 K, R = 0.041 and $R_W = 0.048$ for 1202 unique reflections with $F_o > 2\sigma(F_o)$. A crystal of dimensions $0.1 \times 0.1 \times 0.7$ mm was used. Intensities of 1348 unique reflections were measured to $2\theta_{max} = 120^{\circ}$ in the range $0 \le h \le 30$, $0 \le k \le 10$, $0 \le l \le 6$, with ω scan width of $0.9^{\circ} + 0.15^{\circ}$ tan θ , a scan speed of 3° min⁻¹, and a background counting time of 6 s. The weighting scheme used in the final stage of refinement was w = $[\sigma(F_o)^2 + 0.024F_o]^{-1}$. The residual electron densities in the final difference Fourier map ranged from -0.18 to 0.17 e Å⁻³. Atomic coordinates, temperature factors, and bond lengths and angles have been deposited at the Cambridge Crystallographic Data Centre.

(R)-MTPA Ester 8a.—To a solution of the alcohol 7a (2.5 mg, 11 µmol), prepared as described previously,³ in pyridine (5.0 mm^3) was added (+)-MTPACl (10 mm³, 53 µmol). After 15 h at room temperature, N,N-dimethylpropane-1,3-diamine (6.3 mm³) was added to the reaction mixture. After 10 min, the reaction mixture was diluted with diethyl ether (10 cm^3) and the solution was washed successively with brine, aq. copper(II) sulfate, brine, aq. NaHCO₃ and brine. The solution was dried over Na₂SO₄, and filtered and concentrated under reduced pressure. The residue was subjected to TLC [silica gel; acetonebenzene (1:9)] and then to HPLC (COSMOSIL 5C18-AR, 4.6 \times 150 mm; 55% aq. MeOH) to give (R)-MTPA ester 8a as an oil (3.1 mg, 65%); $\delta_{\rm H}$ 0.761 (3 H, t, J 7.6, 8-CH₂Me), 1.126 (3 H, s, 8-Me), 1.495 (1 H, dq, J 14.2 and 7.6, 8-CH₂Me), 1.716 (1 H, dq, J 14.2 and 7.6, 8-CH₂Me), 1.795 (1 H, m, 6-H), 1.854 (3 H, s, 3-Me), 1.953 (1 H, m, 6-H), 2.200 (3 H, s, 2-Me)₃ 2.385

(1 H, ddd, J 6.4, 8.3 and 18.1, 5-H), 2.457 (1 H, ddd, J 5.5, 6.5 and 18.1, 5-H), 3.421 (3 H, br s, OMe), 5.229 (1 H, dd, J 3.4 and 9.5, 7-H), 7.38 (3 H, m, Ph) and 7.46 (2 H, m, Ph); EI-MS m/z 452 (M⁺, 51%), 422 (8), 219 (100), 191 (22) and 189 (87).

(S)-*MTPA Ester* **8b**.—This compound was obtained in 65% yield from alcohol **7a** with (–)-MTPACl by essentially the same procedure as for the preparation of compound **8a**; $\delta_{\rm H}$ 0.710 (3 H, t, J 7.3, 8-CH₂Me), 1.048 (3 H, s, 8-Me), 1.375 (1 H, dq, J 14.2 and 7.3, 8-CH₂Me), 1.658 (1 H, dq, J 14.2 and 7.3, 8-CH₂Me), 1.854 (3 H, s), 1.866 (1 H, m, 6-H), 2.187 (3 H, s), 2.020 (1 H, m, 6-H), 2.401 (1 H, ddd, J 17.6, 9.2 and 6.4, 5-H), 2.582 (1 H, ddd, J 17.6, 5.9 and 4.8, 5-H), 3.484 (3 H, br s, OMe), 5.253 (1 H, dd, J 3.4 and 9.7, 7-H), 7.355 (3 H, m, Ph) and 7.475 (2 H, m, Ph); EI-MS *m*/*z* 452 (M⁺, 50%), 422 (6), 219 (100), 191 (21) and 189 (89).

Compound 9.—A solution of methyl spicifernin (Me ester of 5b, 55 mg, 0.18 mmol), obtained as previously described,¹ mchloroperbenzoic acid (MCPBA) (121 mg, 0.70 mmol) and NaH_2PO_4 (93 mg, 0.78 mmol) in dichloromethane (2.0 cm³) was stirred for 96 h at 45 °C. The reaction mixture was diluted with dichloromethane (13 cm³) and the solution was washed successively with aq. sodium thiosulfate, aq. NaHCO₃ and brine. After removal of solvent the residue was purified with TLC [silica gel; EtOAc-hexane-AcOH (20:80:1), triple development] to yield compound 9 as an oil (23 mg, 40%); $[\alpha]_{D}^{24}$ ~0 (c 1.0, EtOH); $\delta_{\rm H}$ 0.84 (3 H, t, J 7.3, 9-H₃), 1.20 (3 H, d, J 6.4, 3-Me), 1.33 (3 H, s, 7-Me), 1.86 (1 H, dq, J 14.6 and 7.3, 8-H), 2.00 (1 H, dq, J 14.6 and 7.3, 8-H), 2.01 (3 H, s, 1-H₃), 2.64 (1 H, dd, J 3.9 and 18.1, 5-H), 3.00 (1 H, dd, J 9.3 and 18.1, 5-H), 3.22 (1 H, ddd, J 3.9, 4.9 and 9.3, 4-H), 3.69 (3 H, s, OMe), 3.73 (3 H, s, OMe) and 5.16 (1 H, dq, J 4.9 and 6.4, 3-H); $\delta_{\rm C}$ 8.5, 17.1, 18.2, 21.0, 28.0, 35.8, 44.5, 51.9, 52.3, 59.9, 69.6, 169.9, 172.1, 173.3 and 205.6; FAB-MS m/z 339 (M⁺ + Na, 6%), 317 $(M^+ + H, 26), 301 (24), 257 (99), 225 (14), 197 (36), 154 (100),$ 137 (96), 136 (90), 107 (29), 89 (25) and 59 (12).

Alcohol 10.—A solution of compound 9 (37 mg, 0.12 mmol) and Na_2CO_3 (48 mg, 0.45 mmol) in methanol (1.8 cm³) was stirred for 48 h at room temperature. The reaction mixture was filtered through Celite and diluted with water (10 cm³). The solution was adjusted to pH 2.0 and passed through a Sep Pak C18 Cartridge (Waters Associates). The cartridge was washed with dil. HCl (20 cm³) and was then eluted with methanol (20 cm³). The methanolic solution was concentrated and treated with ethereal diazomethane until a yellow colour persisted. The residue obtained after removal of solvent was purified with TLC [silica gel; EtOAc-hexane-AcOH (30:70:1), triple development] to yield alcohol 10 as an oil (16 mg, 49%); $[\alpha]_{D}^{24} - 6.0$ $(c 1.0, EtOH); \delta_{H} 0.82 (3 H, t, J 7.5, 9-H_3), 1.19 (3 H, d, J 6.4, 3-$ Me), 1.33 (3 H, s, 7-Me), 1.84 (1 H, dq, J 14.5 and 7.5, 8-H), 1.97 (1 H, dq, J 14.4 and 7.4, 8-H), 2.80–3.01 (2 H, m, 5-H₂), 3.71 (3 H, s, OMe), 3.71 (1 H, m, 4-H), 3.72 (3 H, s, OMe) and 3.95 $(1 \text{ H}, \text{m}, 3\text{-}\text{H}); \delta_{C} 8.5, 18.3, 21.0, 27.9, 37.2, 46.8, 51.8, 52.4, 59.9,$ 67.6, 173.3, 174.1 and 206.1; FAB-MS m/z 297 (M⁺ + Na, 85%), 257 (100), 243 (3), 197 (18), 185 (10), 141 (6), 115 (40) and 93 (18).

(R)-*MTPA Ester* 11a.—A solution of alcohol 10 (3.0 mg, 11 μ mol) and (+)-MTPACl (4.2 mm³, 22 μ mol) in pyridine (37 mm³) was kept for 50 h at room temperature. *N*,*N*-Dimethylpropane-1,3-diamine (4.0 mm³) was added to the reaction mixture. After 10 min, the solution was diluted with diethyl ether (2.0 cm³) and washed successively with brine, aq. copper(II) sulfate, brine, aq. NaHCO₃ and brine. After drying over Na₂SO₄, filtration and concentration of the solution, the residue was subjected to TLC [silica gel; hexane–EtOAc–AcOH

(30: 70: 1)] to give (*R*)-MTPA ester **11a** as an oil (3.4 mg, 63%); $\delta_{\rm H}$ 0.826 (3 H, t, J 7.4, 9-H₃), 1.280 (3 H, d, J 7.8, 3-Me), 1.294 (3 H, s, 7-Me), 1.825 (1 H, dq, J 14.2 and 7.4, 8-H), 1.960 (1 H, dq, J 14.2 and 7.4, 8-H), 2.592 (1 H, dd, J 3.4 and 18.1, 5-H), 3.025 (1 H, dd, J 9.8 and 18.1, 5-H), 3.282 (1 H, ddd, J 3.4, 5.8 and 9.8, 4-H), 3.494 (3 H, br s, OMe), 3.660 (6 H, s, 2 × OMe), 5.410 (1 H, dq, J 5.8 and 7.8, 3-H) and 7.38–7.50 (5 H, m, Ph); FAB-MS *m*/*z* 491 (M⁺ + H, 6%), 257 (100), 225 (14), 197 (23) and 189 (45).

(S)-*MTPA Ester* **11b**.—This compound was obtained in 98% yield from alcohol **10** with (-)-MTPACl by essentially the same procedure as for the preparation of compound **11a**; $\delta_{\rm H}$ 0.811 (3 H, t, J 7.3, 9-H₃), 1.267 (3 H, s, 7-Me), 1.346 (3 H, d, J 6.4, 3-Me), 1.781 (1 H, dq, J 14.2 and 7.3, 8-H), 1.936 (1 H, dq, J 14.2 and 7.3, 8-H), 2.475 (1 H, dd, J 18.1 and 3.4, 5-H), 2.961 (1 H, dd, J 18.1 and 10.3, 5-H), 3.254 (1 H, ddd, J 3.4, 5.4 and 10.3, 4-H), 3.527 (3 H, br s, OMe), 3.602 (3 H, s, OMe), 3.680 (3 H, s, OMe), 5.396 (1 H, dq, J 5.4 and 6.4, 3-H) and 7.37–7.50 (5 H, m, Ph); FAB-MS *m*/*z* 491 (M⁺ + H, 14%), 257 (100), 225 (12), 197 (24) and 189 (49).

Acknowledgements

We are grateful to Mr. Katsushi Toyooka, K.I. Chemical Research Institute Co., Ltd, for measuring MS spectra, to Dr. Masakatsu Ichinoe, National Institute of Hygienic Science, Tokyo, for providing the fungus, and to Dr. Hisakazu Yamane, Faculty of Agriculture, The University of Tokyo, for measuring CD spectra. We also thank Professor Yukiteru Katsube and Dr. Mamoru Sato for their kind help in running SHELX 86 in a PC-9801 computer, and the staff of the Research Center for Protein Engineering, Institute for Protein Research, Osaka University for the use of the four-circle diffractometer and the ACOS S-3700/10.

References

- (a) Spiciferone A: H. Nakajima, T. Hamasaki and Y. Kimura, Agric. Biol. Chem., 1989, 53, 2297; (b) Spicifernin: H. Nakajima, T. Hamasaki, S. Maeta, Y. Kimura and Y. Takeuchi, Phytochemistry, 1990, 29, 1739; (c) Spiciferones B and C: H. Nakajima, T. Hamasaki, M. Kohno and Y. Kimura, Phytochemistry, 1991, 30, 2563; (d) Spiciferinone: H. Nakajima, Y. Kimura and T. Hamasaki, Phytochemistry, 1992, 31, 105.
- 2 H. Nakajima, R. Matsumoto, Y. Kimura and T. Hamasaki, J. Chem. Soc., Chem. Commun., 1992, 1654; H. Nakajima, H. Fujimoto, R. Matsumoto and T. Hamasaki, J. Org. Chem., 1993, 58, 4526.
- 3 H. Nakajima, H. Fujimoto, Y. Kimura and T. Hamasaki, Biosci. Biotechnol. Biochem., 1993, 57, 1938.
- 4 J. A. Dale and H. S. Mosher, J. Am. Chem. Soc., 1973, 95, 512;
 I. Ohtani, T. Kusumi, Y. Kashman and H. Kakisawa, J. Am. Chem. Soc., 1991, 113, 4092; J. Org. Chem., 1991, 56, 1296.
- 5 M. Ubukata, X.-C. Cheng, M. Isobe and K. Isono, J. Chem. Soc., Perkin Trans. 1, 1993, 617.
- 6 G. M. Sheldrick, SHELX 86, Crystallographic Computing 3, ed. G. M. Sheldrick, C. Krüger and R. Goddard, Oxford University Press, 1985, pp. 175–189.
- 7 T. Ashida, HBLS-V, The Universal Crystallographic Computing System, Computation Center, Osaka University, 1973, p. 55.
- 8 International Tables for X-Ray Crystallography, Kynoch Press, Birmingham, 1974.
- 9 C. K. Johnson, ORTEP-II Oak Ridge National Laboratory, Oak Ridge, TN, 1976.

Paper 3/07355F Received 14th December 1993 Accepted 24th February 1994